

ARACHIDONATE EPOXYGENASE: INHIBITORS AND METABOLITE ANALOGUES

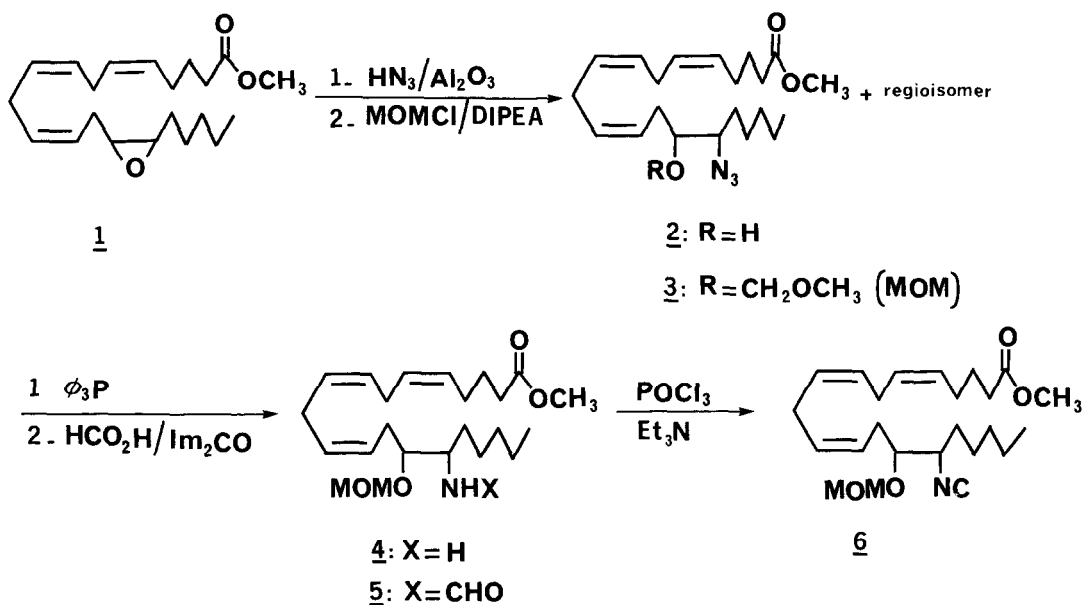
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Summary: The preparation and initial *in vitro* evaluation of a series of potent arachidonate epoxygenase inhibitors and heteroatom analogues of epoxyeicosatrienoic acid are described.

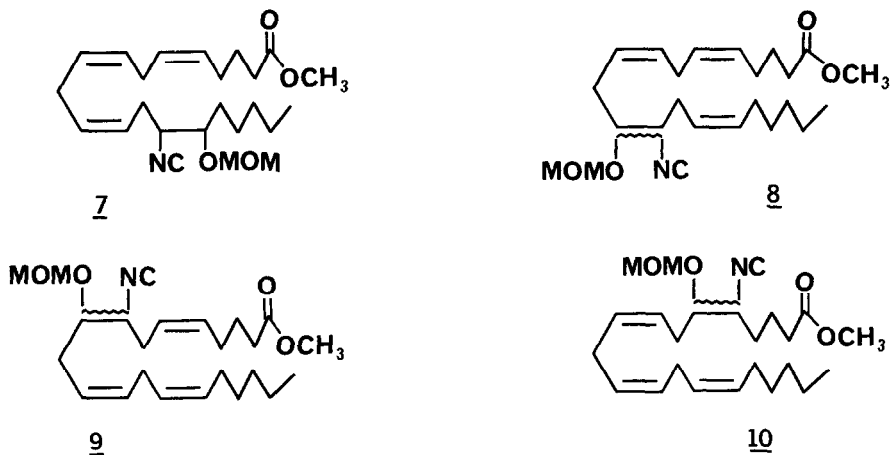
Recently, evidence has been presented for a novel mode of eicosanoid production mediated by cytochrome P-450 and requiring NADPH¹. Designated the epoxygenase pathway, this route produces four regioisomeric epoxyeicosatrienoic acids (EETs). The EETs exhibit significant *in vitro* biological activity² and have been detected in mammalian tissue³ and human urine⁴. In recognition of the need⁵ to selectively intervene in epoxygenase metabolism and to unravel the physiological role of its metabolites, we report herein the preparation and *in vitro* evaluation of a series of potent epoxygenase inhibitors and EET heteroatom analogues⁶.

Exposure of methyl 14,15-EET 1⁷ to anhydrous hydrazoic acid (Et₂O/PhH 5:1, 10 min) in the presence of neutral alumina (Woelm 200) according to the general procedure of Posner and Rogers⁸ furnished (Scheme I) a chromatographically separable mixture of azidohydrin 2^{9,10} and its positional isomer (78%, 1.7:1 ratio). TLC: SiO₂, Et₂O/hexane 1:2, R_f ~ 0.26 and 0.28, respectively. Treatment of 2 with diisopropylethylamine (5 equiv) and chloromethyl methyl ether (4.8 equiv) for 60 h in dry dichloromethane gave methoxymethyl (MOM) ether 3 (90%) after extractive isolation and chromatography (SiO₂, Et₂O/hexane 2:1, R_f ~ 0.5). Selective azide reduction¹¹ by triphenylphosphine (2 equiv, THF, 24 h), hydrolysis of the intermediate iminophosphorane (THF/H₂O 7:1, 36 h), and purification (SiO₂, 10% MeOH/CH₂Cl₂, R_f ~ 0.29) afforded amine 4 (86%) which was directly formylated¹² using carbonyldiimidazole (Im₂CO) and formic acid in THF for 1 h. The resultant formamide 5 (SiO₂:Et₂O, R_f ~ 0.46) was dehydrated with POCl₃ (5 equiv) and Et₃N (20 equiv) in Et₂O/Petroleum ether (1:1) at 40°C for 1 h. Addition of the reaction mixture to ice cold 5% aqueous NaHCO₃, extractive isolation, and chromatography (SiO₂, Et₂O/hexane 1:1, R_f ~ 0.45) provided isonitrile 6 in 85% yield from 4.

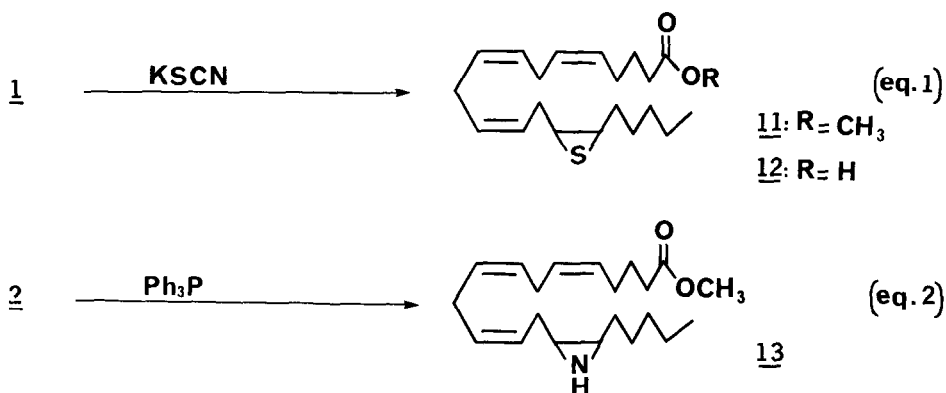


Scheme I

Repetition of the above sequence using the positional isomer of azidohydrin 2 afforded 7 in 67% overall yield. Isonitriles 8-10 were prepared analogously from the corresponding methyl EETs⁷. In the case of the latter compounds, however, no attempt was made to separate regioisomers. Prior to testing, 6-10 were saponified (NaOH, 10 h) in THF/H₂O (3:1) and the sodium salts isolated using BioRad SM-2 resin¹³.



Physico-chemical considerations led to the working hypothesis that isonitriles 6-10 could function as powerful site specific inactivators of epoxygenase activity by coordination to the heme iron of cytochrome P-450¹⁴. Indeed, analysis of the isonitrile bound cytochrome P-450 difference spectrum of 6-10 revealed high binding affinity. Over a range of 0.1-100 μM 6-10, the spectral dissociation constant utilizing 3 μM hemoprotein was $\sim 7 \mu\text{M}$. The metabolism of 100 μM arachidonic acid by liver microsomal fractions from phenobarbital induced rats was inhibited 75% by 6-10 at 5 μM . Significantly, arachidonate metabolism by PG synthetase purified from ram seminal vesicles, soybean lipoxygenase and by sonicated human platelets was unaffected by 6-10 at 5 μM ; 100 μM 6-10 blocked metabolism in these systems by no more than 20%.



It was also deemed desirable to examine EET heteroatom analogues for epoxygenase inhibition. Treatment of 1 with excess potassium thiocyanate¹⁶ in MeOH (65°C, 36 h) under argon (eq. 1), acidification (pH 4), and extractive isolation evolved *cis*-episulfides 11 (55%) and 12 (27%) (SiO₂, Et₂O/Hexane 1:2, R_f \sim 0.55 and 0.24, respectively). When 2 was allowed to interact with triphenylphosphine (1 equiv, 18 h) in dry benzene (eq. 2), *cis*-aziridine 13^{6b} was obtained (SiO₂, CH₃CN, R_f \sim 0.14). Testing of the respective sodium salts showed 13 was comparable to the isonitriles in inhibiting arachidonate metabolism by liver microsomal fractions. In contrast, episulfide 12 was an order of magnitude less potent. Studies are in progress to assess the EET agonist/antagonist activity of EET heteroatom analogues.

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References and Notes

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9. Satisfactory spectral data were obtained for all new compounds using chromatographically homogeneous samples.
10. Physical data. 2: NMR (CDCl₃, 90 MHz) δ 0.88 (3H,t), 1.08-2.52 (17H,m), 2.56-2.96 (4H,m), 3.04-3.60 (2H,m), 3.64 (3H,s), 5.12-5.64 (6H,m); IR (CHCl₃): 3580, 2100, 1730 cm⁻¹. 3: NMR (δ) 0.88 (3H,t), 1.08-2.48 (16H,m), 2.64-2.92 (4H,m), 3.08-3.64 (2H,m), 3.36 (3H,s), 3.64 (3H,s), 4.52-4.80 (2H,m), 5.12-5.52 (6H,m); IR (CHCl₃): 2120, 1735 cm⁻¹. 4: NMR (δ) 0.88 (3H,t), 1.08-2.44 (18H,m), 2.52-2.92 (5H,m), 3.20-3.48 (1H,m), 3.36 (3H,s), 3.64 (3H,s), 4.52-4.76 (2H,m), 5.08-5.52 (6H,m). 5: NMR (δ) 0.88 (3H,t), 1.08-2.44 (16H,m), 2.52-2.88 (4H,m), 3.36 (3H,s), 3.64 (3H,s), 3.88-4.20 (1H,m), 4.48-4.76 (2H,m), 5.12-5.50 (6H,m), 8.12-8.24 (1H,m); IR (CHCl₃) 1730, 1690 cm⁻¹. 6: NMR (δ) 0.88 (3H,t), 1.08-2.52 (16H,m), 2.64-2.96 (4H,m), 3.36 (3H,s), 3.64 (3H,s), 3.40-3.72 (2H,m), 4.52-4.80 (2H,m), 5.08-5.64 (6H,m); IR (CHCl₃) 2150, 1735 cm⁻¹. 11: NMR (δ) 0.88 (3H,t), 1.26-1.84 (10H,m), 2.06 (3H,t), 2.24-2.53 (4H,m), 2.70-3.00 (6H,m), 3.64 (3H,s), 5.27-5.94 (6H,m); PCI mass spec m/e: 351, 319, 287, 269. 13: NMR (δ) 0.88 (3H,t), 1.12-2.40 (19H,m), 2.60-2.92 (4H,m), 3.64 (3H,s), 5.12-5.52 (6H,m); mass spec m/e: 332, 318, 302, 262.
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